

## Results of the Toxoplasmosis Screening in 9311 Pregnant Women in a Tertiary Center in Turkey

### Türkiye'de Üçüncü Basamak Bir Merkezde Yapılan 9311 Gebede Toksoplazma Taramasının Sonuçları

Özgür KOÇAK<sup>1</sup>([iD](#)), Özgür KAN<sup>1</sup>([iD](#))

<sup>1</sup> Department of Gynecology and Obstetrics, Faculty of Medicine, Hitit University, Çorum, Turkey

**Cite this article as:** Koçak Ö, Kan Ö. Results of the toxoplasmosis screening in 9311 pregnant women in a tertiary center in Turkey. FLORA 2020;25(3):332-8.

#### ABSTRACT

**Introduction:** *Toxoplasma gondii* is an important parasite that can cause permanent sequelae to the fetus when infected during pregnancy in humans. Although the frequency of this parasite varies widely between countries, it is known that it is common in our country. The aim of this study was to determine the seroprevalence of toxoplasma in pregnant women admitted to a tertiary hospital in central Anatolia and to evaluate the pregnancy outcomes together with seroprevalence.

**Materials and Methods:** A total of 9311 patients admitted to a tertiary hospital between January 2016 and December 2018 were included into the study. After serological examination, avidity test was performed in cases suggestive of acute infection. Amniocentesis was recommended to be performed by Polymerase Chain Reaction (PCR) in patients with low avidity.

**Results:** The frequency of *Toxoplasma immunoglobulin (Ig) G* and *M* seropositivity rates were 20.3% and 0.28%, respectively. Low avidity was found in approximately 27% of the patients with IgM positivity, and only 15.4% had low avidity by confirmatory test. One patient could not be reached during follow-up. PCR was performed in 4 patients whose low avidity value was confirmed by re-tests and all of their PCR results were reported negative. No cases of congenital toxoplasmosis were detected during the 3 years in our clinic.

**Conclusion:** The inclusion of toxoplasma in routine screening programme is still controversial and differs between countries. Screening in areas with a high rate of toxoplasma, such as in our country, may be rational. If infection is detected, treatment may be recommended because it may reduce the transmission to the fetus.

**Key Words:** *Toxoplasma*; Screening; Pregnancy; Antenatal; Turkey

## ÖZ

## Türkiye'de Üçüncü Basamak Bir Merkezde Yapılan 9311 Gebede Toksoplazma Taramasının Sonuçları

Özgür KOÇAK<sup>1</sup>, Özgür KAN<sup>1</sup><sup>1</sup> Hitit Üniversitesi, Kadın Hastalıkları ve Doğum Anabilim Dalı, Çorum, Türkiye

**Giriş:** Toksoplazma gondii, insanlarda hamilelik sırasında enfekte olduğunda fetusa kalıcı sekellere neden olabilecek önemli bir parazittir. Bu parazitin sıklığı ülkeler arasında büyük farklılıklar gösterse de ülkemizde yaygın olduğu bilinmektedir. Bu çalışmanın amacı, Orta Anadolu'daki tersiyer merkez hastaneye başvuran gebe kadınlarda toksoplazmanın seroprevalansını belirlemek ve gebelik sonuçlarını seroprevalansı ile birlikte değerlendirmektir.

**Materyal ve Metod:** Çalışmaya Ocak 2016-Aralık 2018 tarihleri arasında hastaneye başvuran toplam 9311 hasta alındı. Serolojik incelemeden sonra akut enfeksiyonu düşündürülen olgulara avidite testi yapıldı. Düşük avidite saptananlarda Polimeraz Zincir Reaksiyonu (PCR) yapılması için amniyosentez önerildi.

**Bulgular:** Toksoplazma immünglobulin (Ig) G ve M seropozitiflik sıklığı sırasıyla %20.3 ve %0.28 idi. IgM pozitifliği olan hastaların yaklaşık %27'sinde düşük avidite bulunmuştur ve doğrulama testiyle sadece %15.4'ünde düşük avidite saptanmıştır. Düşük avidite değeri tekrar testlerle doğrulanan 4 hastaya PCR yapıldı ve tüm PCR sonuçları negatif olarak bildirildi. Kliniğimizde 3 yıl boyunca hiçbir hastada konjenital toksoplazmoz olgusu saptanmadı.

**Sonuç:** Rutin tarama programına toksoplazmanın dahil edilmesi halen tartışmalıdır ve ülkeler arasında farklılık göstermektedir. Ülkemizde olduğu gibi yüksek oranda toksoplazma bulunan bölgelerde tarama yapılması rasyonel olabilir. Enfeksiyon tespit edilirse, fetusa geçişi azaltılabileceğinden tedavi önerilebilir.

**Anahtar Kelimeler:** Toksoplazma; Tarama; Gebelik; Antenatal; Türkiye

## INTRODUCTION

*Toxoplasma gondii* is a protozoan parasite generating a parasitic disease "toxoplasmosis" in humans and in animals<sup>[1]</sup>. In adults, toxoplasmosis may be asymptomatic<sup>[2]</sup>. If symptomatic, the only symptoms may be flu-like; headache, muscle aches, etc. The infection is usually acquired during adolescence and childhood<sup>[3]</sup>. It may cause serious problems if infection occurs during pregnancy and if transmitted to the fetus. In this case, it is called congenital toxoplasmosis, and the incidence of infection during pregnancy ranges from 1-8 per 1000 pregnancies<sup>[4]</sup>. Fetal infection occurs with the transmission of the parasite through the placenta<sup>[5]</sup>. Gestational week of the pregnancy impact the clinical features. Intracranial calcifications, hydrocephalus, echogenic bowel, hepatomegaly, splenomegaly, intrahepatic calcifications, intrauterine growth restriction, ascites, pericardial and/or pleural effusions, hydrops fetalis, fetal demise, placental densities may be the sonographic findings of congenital toxoplasmosis<sup>[6]</sup>. All infections, even all fetal findings, do not lead to serious

sequelae. For that reason, it is very important for clinicians to detect the infected pregnant woman.

The aim of this study was to retrospectively evaluate the *Toxoplasma* IgG and IgM positive pregnant woman, and thus to determine the seroprevalence and detect the postnatal results of these pregnancies in a tertiary center.

## MATERIALS AND METHODS

This is a retrospective study, and all the data were obtained from the hospital database, records, and if necessary, by phone calls. The study was conducted at a tertiary center. Approval of the local ethics committee was received before the study started (Date: April, 25, 2019; No: 2019-145). Female patients who applied for pregnancy follow-up in our obstetrics outpatient clinic between January 2016 and December 2018 were included into this study.

A total of 9311 women with proved intrauterine pregnancy by ultrasound at the first trimester were included into this study. Demographical features including age, obstetrical history

and comorbidities at the first prenatal visit were reviewed from patient files.

*Toxoplasma* IgG and/or IgM antibodies were tested according to the clinician's prediction or the women's complaints during pregnancy follow-up. Generally, *Toxoplasma* tests are performed when the intrauterine pregnancy is proven, and it is generally done at the first visit in the first trimester.

*Toxoplasma* IgG and *Toxoplasma* IgM were analyzed using Cobas 6000 diagnostic system (Roche Diagnostics Ltd, Switzerland) according to the manufacturer's instructions. The Elecsys Toxo IgM and IgG assay was used for the quantitative determination of specific antibodies to toxoplasmosis in human serum or plasma samples. The tests were performed on the Cobas 6000 Analyzer. According to the manufacturer's instructions, values of IgG and IgM levels greater than 3 IU/mL and 1 IU/mL were regarded as positive, respectively. Pregnant woman who had high levels of *Toxoplasma* IgG and IgM levels were tested with avidity test to determine acute or chronic infection.

While avidity index greater than 0.300 for *Toxoplasma* IgG was regarded as high avidity, less than 0.200 was regarded as low avidity. Low avidity may be an indicator of acute infection. All positive test results were controlled and retested by the same way. Values between 0.200 and 0.300 were considered as intermediate values,

and further investigations were made. Further investigations were done by Polymerase chain reaction (PCR) which is one of the basic tests of modern molecular biology. One DNA molecule can be reproduced millions of times and quickly diagnosed by PCR.

Statistical Package for the Social Sciences (SPSS) version 22.0 was used for statistical calculations. Data were presented with the number of pregnant women and percentage and defined by median (minimum–maximum). Numbers and percentages were used as descriptive statistical methods in evaluating the data.

## RESULTS

Totally, 9311 pregnant women were included into this study. Median age of the population was 25 years (range: 16-47 years). *Toxoplasma* IgG positivity frequency varied between 18.5% and 24.5% by the years, and 3 years average IgG positivity was 20.3% (Table 1).

*Toxoplasma* IgM seropositivity ranged between 0.26% and 0.35%. Three years average IgM seropositivity was 0.28% (Table 2).

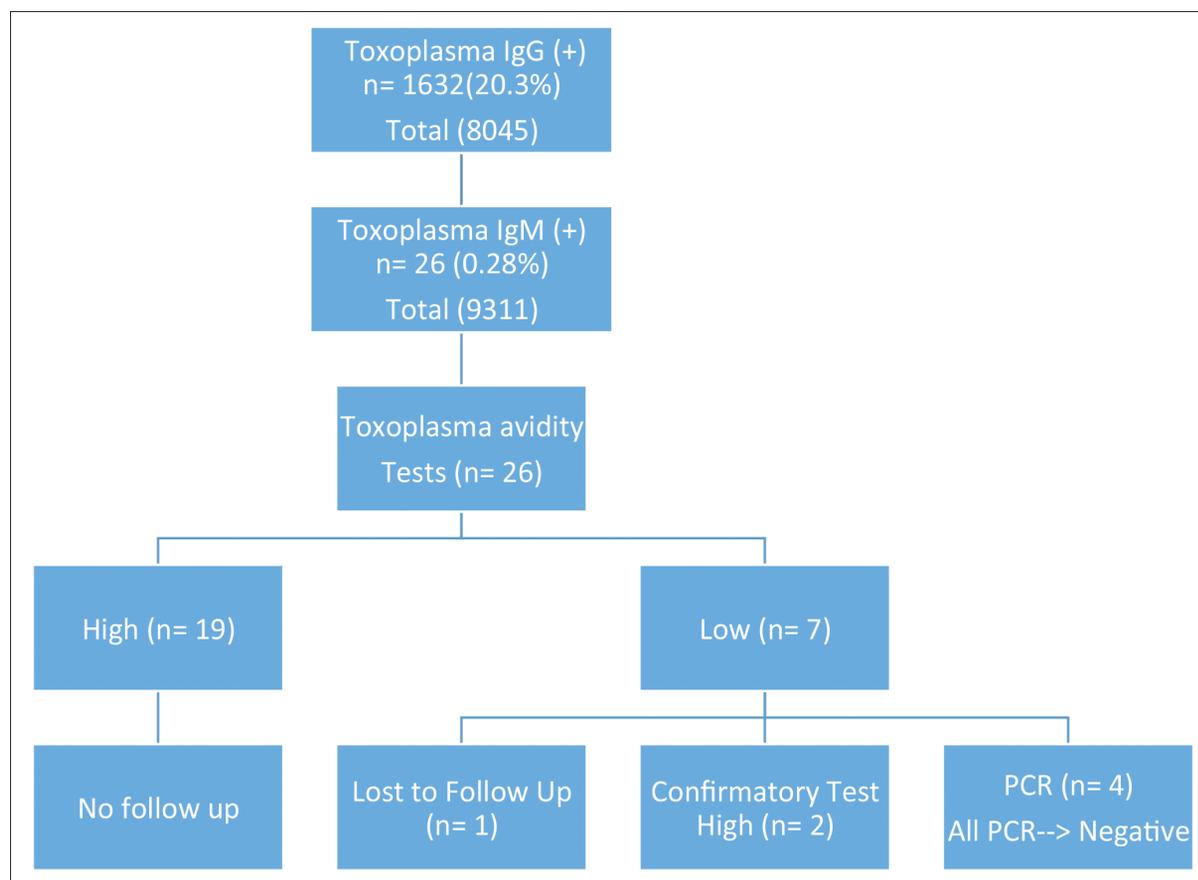
Avidity test was performed in all 26 women who had IgM positivity. Low avidity was detected in seven of those 26 women. Spiramycin was started empirically in those seven women with low avidity. Avidity was checked with the same laboratory two weeks later. One woman was lost to follow up during the procedure. Two weeks

**Table 1. Toxoplasma IgG seropositivity by years**

Years	Total test	Toxoplasma IgG (+) n (%)
2016	1716	421 (24.5%)
2017	2787	518 (18.5%)
2018	3542	693 (19.6%)
Total	8045	1632 (20.3%)

**Table 2. Toxoplasma IgM seropositivity by years**

Years	Total test	Toxoplasma IgM (+) n (%)
2016	1953	7 (0.35%)
2017	3080	8 (0.26%)
2018	4278	11 (0.33%)
Total	9311	26(0.28%)



**Figure 1.** The flow chart of the cases.

later, the avidity test of two women was found to be high in repeated test. Four women with low avidity test underwent PCR after amniocentesis. All women had negative PCR results.

Congenital toxoplasmosis has a triad that consists of intracranial calcifications, chorioretinitis and hydrocephalus. No fetus was detected with this triad at birth. Congenital toxoplasmosis may not show any clinical manifestations on routine physical examination. For that reason, these suspected four fetuses were detailed in central nervous system (CNS) imaging and ophthalmologic examination. At the end of these examinations, no pathologic findings were detected. These fetuses were followed up to 6 months to three years, and no congenital toxoplasmosis was detected in any fetus during the follow up.

## DISCUSSION

*T. gondii* seroprevalence among childbearing-aged women (aged between 15-45 years)

has a range of 8.2%-63.2% in Europe<sup>[7]</sup>. In the United States of America, it is 6.2%<sup>[8]</sup>. The seroprevalence of *T. gondii* has decreased over the past 20 years<sup>[8]</sup>. Toxoplasmosis infection may cause serious complications if occurs during pregnancy<sup>[9]</sup>.

Fetal infection risk increases steeply with gestational age. A meta-analysis has shown that seroconversion rate is 15% at 13 weeks, 44% at 26 weeks and at 36 week, it is as high as 71%<sup>[10]</sup>. Although fetal infection risk increases by gestational age, the risk of clinical sequelae decreases.

The need for routine screening of all pregnant women for toxoplasmosis is controversial because maternal infection during pregnancy is as low as 1-8 to 1000 in UK<sup>[4]</sup>. For this reason, some countries such as the United States, United Kingdom, Canada and some parts of Europe are against screening<sup>[11-13]</sup>. On the other hand, some

countries in Europe, where seroprevalence is higher, perform screening as routine follow up<sup>[14,15]</sup>. Even some studies recommend screening for congenital toxoplasmosis monthly, bimonthly or with three months intervals<sup>[12,15]</sup>.

It is mentioned that some countries are against screening but if there is high clinical suspicion of acute toxoplasmosis infection during pregnancy, diagnostic tests should be performed<sup>[6]</sup>. These high clinical manifestations are fever and adenopathy in woman and intracranial calcification and ventricular dilatation of the fetus.

A review in 2019 investigating toxoplasmosis in Turkey reported that toxoplasmosis seropositivity rate was one third of the population<sup>[16]</sup>.

In our country, the Ministry of Health has no specific policy regarding toxoplasmosis screening during pregnancy. There is no sentence about *Toxoplasma* in the prenatal care management guide published in 2014 by Ministry of Health of Turkey<sup>[17]</sup>. Turkish Perinatology Society does not recommend testing for *Toxoplasma* serology during pregnancy. Therefore, every clinician's approach to *Toxoplasma* is different according to their own knowledge and experience in our country.

In our study, we examined more than 9.000 pregnant women and we found the *Toxoplasma* IgG positivity rate as 20.3% and IgM positivity rate as 0.28% in mid-northern region of Turkey. Ocak et al. have reported anti-*Toxoplasma* IgG antibody as 52.1% and anti-*Toxoplasma* IgM antibody as 0.54% in southern Turkey<sup>[18]</sup>. Harma et al. have found *Toxoplasma* IgG positivity as 60.4% and IgM positivity as 3% in a city in Southeastern Turkey<sup>[19]</sup>. These studies were reported in 2004 and 2009. The prevalence of higher IgM positivity rates in these studies may be due to the year they were published. As mentioned before, *Toxoplasma* IgG and IgM positivity rate has decreased in the past decades due to training, compliance rules of hygiene, and raising awareness<sup>[8]</sup>. This may be the case in Turkey<sup>[20]</sup>.

In this study, 9311 pregnant women in total were tested, and IgM positivity was detected in 26 of them (0.28%). In these women, IgG was also positive. If a woman has both IgM positivity and IgG positivity, Hedman et al. first described

IgG avidity test in 1989 in such cases. In this test, high avidity excludes recent infection and low avidity suggests the contrary<sup>[21]</sup>. The detection of recently infected pregnant women is very crucial due to transmission rates. In a meta-analysis, transmission rate has been confirmed as 15% at 13 weeks, 44% at 26 weeks, and 71% at 36 weeks<sup>[10]</sup>. Although transmission rate increases as the gestational week increases, the sequelae rate decreases. In one study, it has been shown that sequelae rate decreases by approximately 4% at every week<sup>[6]</sup>. As the gestational week will play an important role in fetal infection, factors such as parasite load, strain of the parasite, and mother's immune system play a role in transmission<sup>[6]</sup>.

In a paper reported by Peyron et al., transmission rate has been reported to be about half in women treated three weeks after infection detection compared to the women treated eight weeks after infection<sup>[22]</sup>. Some other studies have also shown that early treatment of toxoplasmosis lowers transmission rates and the sequelae<sup>[23,24]</sup>.

In this study, 26 women who were positive for *Toxoplasma* IgM were recommended to undergo avidity testing. Spiramycin treatment was started empirically to the women with positive *Toxoplasma* IgM. Treatment was discontinued in women with high avidity test and continued with low avidity. Avidity test revealed low in seven women.

Avidity test is used to determine when antibodies are formed<sup>[25]</sup>. If the avidity is high, the infection is thought to have occurred before four months<sup>[26]</sup>. The benefit of low avidity in clinical use is less than high avidity because there are many causes of low avidity. In some cases, low avidity can be detected during months, which creates a dilemma in which gestational infection is occurred. Therefore, looking at low avidity test can be misleading to conclude that there is an acute infection without confirmatory tests<sup>[27]</sup>. In these situations, agglutination assays and/or specific immunoglobulin tests may be used<sup>[28]</sup>.

In this study, 26 women who were positive for *Toxoplasma* IgM were recommended to undergo avidity testing. Avidity test revealed low avidity in seven women that may indicate acute

infection. The avidity test was repeated in six women because one of them missed the follow up, and test results of two of the six women reported high avidity.

If toxoplasmosis is clinically suspicious, amniocentesis and PCR are recommended after 18 weeks to show *T. gondii* DNA in the amniotic fluid<sup>[29]</sup>. In a systematic review, the specificity of PCR in amniotic fluid has been detected as 98% and sensitivity as approximately 74% in all trimesters<sup>[29]</sup>.

However, the most important factor affecting the sensitivity of PCR is the time from the onset of infection. It is known that the virus will reach the amniotic fluid after a certain time of infection. Therefore, *Toxoplasma* DNA cannot always be detected by PCR in every infected women. For this reason, congenital toxoplasmosis diagnosis is based on clinical suspicion and confirmation with the laboratory.

In our study, PCR was performed in four women and all of them were reported as negative.

There is no randomized controlled trial showing that toxoplasmosis treatment during pregnancy improves pregnancy outcomes. However, prenatal treatment is offered to reduce the risk of possible congenital toxoplasmosis<sup>[10]</sup>.

There are two treatment options that reduce congenital toxoplasmosis; spiramycin and pyrimethamine-sulfadiazine plus folinic acid. If the women are < 18 weeks of gestation at the time of diagnosis, spiramycin is the first choice, otherwise pyrimethamine-sulfadiazine plus folinic acid is the choice.

In this study, spiramycin was started in seven women with suspected acute *toxoplasma* infection and no newborn congenital toxoplasmosis was detected. It has a triad that consists of intracranial calcifications, chorioretinitis and hydrocephalus. However, < 10 percent of cases has this triad at birth<sup>[30]</sup>. Congenital *toxoplasma* was not detected in all of these women's babies.

In conclusion, trends in toxoplasmosis screening are different between countries. It is included in routine screening program in countries with high positivity rates such as France. It can

be said that positivity rate is high in our country. Therefore, we think that routine screening will be beneficial and cost effective.

### ETHICS COMMITTEE APPROVAL

Ethics committee approval was obtained for this study from of Hitit University Non-Invasive Research Ethical Committee (Decision No: 2019-145 Date: 26.04.2019)

### CONFLICT of INTEREST

The authors declare that they have no conflict of interest.

### AUTHORSHIP CONTRIBUTIONS

Concept/Design: ÖK, ÖK

Analysis/Interpretation: ÖK, ÖK

Data Acquisition: ÖK, ÖK

Writing: ÖK, ÖK

Final Approval: ÖK, ÖK

### REFERENCES

1. Centers for Disease Control and Prevention (CDC). Parasites - Toxoplasmosis (*Toxoplasma* infection) Erişim tarihi: 23 Ağustos 2015. Available from: <https://www.cdc.gov/parasites/toxoplasmosis/epi.html>
2. Hunter CA, Sibley LD: Modulation of innate immunity by *Toxoplasma gondii* virulence effectors. *Nat Rev Microbiol* 2012;10:766.
3. Welton NJ, Ades AE. A model of toxoplasmosis incidence in the UK: Evidence synthesis and consistency of evidence. *JRSS-C Applied Statistics* 2005;54:385
4. Gilbert RE, Peckham CS. Congenital toxoplasmosis in the United Kingdom: to screen or not to screen? *J Med Screen* 2002;9:135-41.
5. Remington JS MR, Thulliez P, Desmonts G. *Toxoplasmosis. In: Remington KJ, Wilson CB, Baker CJ (eds). Infectious disease of the fetus and newborn infant. 6<sup>th</sup> ed. Philadelphia: Elsevier Saunders, 2006:947.*
6. Maldonado YA, Read JS. Diagnosis, Treatment, and Prevention of Congenital Toxoplasmosis in the United States. *Pediatrics* 2017;139:e20163860
7. Pappas G, Roussos N, Falagas ME. *Toxoplasmosis snapshots: global status of Toxoplasma gondii seroprevalence and implications for pregnancy and congenital toxoplasmosis. Int J Parasitol* 2009;39:1385-94.
8. Jones JL, Kruszon-Moran D, Rivera HN, Price C, Wilkins PP. *Toxoplasma gondii* seroprevalence in the United States 2009-2010 and comparison with the past two decades. *Am J Trop Med Hyg* 2014;90:1135-9.

9. Hohlfeld P, Daffos F, Thulliez P, Aufrant C, Couvreur J, MacAleese J, et al. Fetal toxoplasmosis: outcome of pregnancy and infant follow-up after in utero treatment. *J Pediatr* 1989;115:765-9.
10. Thiebaut R, Leproust S, Chene G, Gilbert R. Effectiveness of prenatal treatment for congenital toxoplasmosis: a meta-analysis of individual patients' data. *Lancet* 2007;369:115-22.
11. Stein IF, Leventhal ML. Amenorrhea associated with bilateral polycystic ovaries. *Am J Obstet Gynecol* 1935;29:181-91.
12. Paquet C, Yudin MH. Toxoplasmosis in pregnancy: prevention, screening, and treatment. *J Obstet Gynaecol Can* 2018;40:e687-93.
13. Practice bulletin no. 151: Cytomegalovirus, parvovirus B19, varicella zoster, and toxoplasmosis in pregnancy. *Obstet and Gynecol* 2015;125:1510-25.
14. Gilbert R, Gras L. Effect of timing and type of treatment on the risk of mother to child transmission of *Toxoplasma gondii*. *BJOG* 2003;110:112-20.
15. Wallon M, Peyron F, Cornu C, Vinault S, Abrahamowicz M, Kopp CB, et al. Congenital toxoplasma infection: monthly prenatal screening decreases transmission rate and improves clinical outcome at age 3 years. *Clin Infect Dis* 2013;56(9):1223-31.
16. Koloren Z, Dubey JP. A review of toxoplasmosis in humans and animals in Turkey. *Parasitology* 2019;147:1-17.
17. *Prenatal Care Management Guide*, 2014.
18. Ocak S, Zeteroglu S, Ozer C, Dolapcioglu K, Gungoren A. Seroprevalence of *Toxoplasma gondii*, rubella and cytomegalovirus among pregnant women in southern Turkey. *Scandinavian J Infect Dis* 2007;39:231-4.
19. Harma M, Harma M, Gungen N, Demir N. Toxoplasmosis in pregnant women in Sanliurfa, Southeastern Anatolia City, Turkey. *J Egypt Soc Parasitol* 2004;34:519-25.
20. İnci M, Yağmur G, Aksebzeci T, Kaya E, Yazar S. Kayseri'de kadınlarda *Toxoplasma gondii* seropozitifliğinin araştırılması. *Türkiye Parazitol Derg* 2009;33:191-4.
21. Hedman K, Lappalainen M, Seppä I, Mäkelä O. Recent primary toxoplasma infection indicated by a low avidity of specific IgG. *J Infect Dis* 1989;159:736-40.
22. Peyron F, McLeod R, Ajzenberg D, Contopoulos-Ioannidis D, Kieffer F, Mandelbrot L, et al. Congenital toxoplasmosis in France and the United States: One Parasite, Two Diverging Approaches. *PLoS Negl Trop Dis* 2017;11:e0005222.
23. Sickinger E, Gay-Andrieu F, Jonas G, Schultess J, Stieler M, Smith D, et al. Performance characteristics of the new ARCHITECT toxo IgG and Toxo IgG avidity assays. *Diagn Microbiol Infect Dis* 2008;62(3):235-44.
24. Jost C, Touafek F, Fekkar A, Courtin R, Ribeiro M, Mazier D, et al. Utility of Immunoblotting for early diagnosis of toxoplasmosis seroconversion in pregnant women. *Clin Vaccine Immunol* 2011;18:1908-12.
25. Bobic B, Klun I, Vujanic M, Nikolic A, Ivovic V, Zivkovic T, et al. Comparative evaluation of three commercial *Toxoplasma*-specific IgG antibody avidity tests and significance in different clinical settings. *J Med Microbiol* 2009;58:358-64.
26. Beghetto E, Buffolano W, Spadoni A, Del Pezzo M, Di Cristina M, Minenkova O, et al. Use of an immunoglobulin G avidity assay based on recombinant antigens for diagnosis of primary *Toxoplasma gondii* infection during pregnancy. *J Clin Microbiol* 2003;41(12):5414-8.
27. Villard O, Breit L, Cimon B, Franck J, Fricker-Hidalgo H, Godineau N, et al. Comparison of four commercially available avidity tests for *Toxoplasma gondii*-specific IgG antibodies. *Clin Vaccine Immunol* 2013;20:197-204.
28. Robert-Gangneux F, Darde ML. Epidemiology of and diagnostic strategies for toxoplasmosis. *Clin Microbiol Rev* 2012;25:264-96.
29. de Oliveira Azevedo CT, do Brasil PE, Guida L, Lopes Moreira ME. Performance of polymerase chain reaction analysis of the amniotic fluid of pregnant women for diagnosis of congenital toxoplasmosis: A systematic review and meta-analysis. *PLoS ONE* 2016;11:e0149938.
30. Tamma P. Toxoplasmosis. *Pediatr Rev* 2007;28:470-1.

#### Address for Correspondence/Yazışma Adresi

Dr. Öğr. Üyesi Özgür KOÇAK  
Hitit Üniversitesi Tıp Fakültesi,  
Kadın Hastalıkları ve Doğum Anabilim Dalı,  
Çorum-Türkiye  
E-mail: dr.ozgur@hotmail.com